

PULSE ELECTRIC FIELD EFFECT ON CERVICAL CANCER CELL:
A PLANT BASED ANTI-CANCER STUDY

SUHASSNI GANESON

A thesis submitted in
fulfillment of the requirement for the award of the
Degree of Master of Electrical Engineering



PTTAUTHM
PERPUSTAKAAN TUNKU TUN AMINAH

Faculty of Electrical and Electronic Engineering
Universiti Tun Hussein Onn Malaysia

NOVEMBER 2018

For my father Ganeson Sengan, my mother Saraswathy Arjunan, my siblings Hemaraaj Ganeson, Devaraaj Ganeson, Thurgashini Ganeson, my husband Dayamugil Ponnudurai, my uncle Vasantha Rajan and my aunt Khaitiri Devi.



PTTA UTHM
PERPUSTAKAAN TUNKU TUN AMINAH

ACKNOWLEDGEMENT

Foremost, I would like to express my heartfelt gratitude to the Almighty God for given me the life, health, wealth and wisdom has bestowed upon me during this research, and indeed, throughout my life.

My deepest appreciation and profound gratitude goes to my supervisor, Associate Professor Muhammad Mahadi Abdul Jamil for his unwavering guidance, enthusiastic encouragement and mentorship in all the time of research and writing of this thesis. I could not have imagined having a better supervisor. Without his encouragement and patience, I would have not completed the course of Master research. I would like to thank my co-supervisor Dr. Muhammad Nazib Bin Adon for his support and guidance throughout the course of my research work.

I extend my thanks to Ministry of Highest Education for the financial support throughout my studies through MyBrain15 (MyMaster) scholarship as well as Universiti Tun Hussein Onn Malaysia through Contract Grant U551. Finally, special thanks to my beloved family, especially my aunty, uncle and my husband, for their unceasing moral support and precious love, have empowered me to accomplish my thesis. Not forgetting to all my friends, especially Nisha Sunthra for all the help and the good time shared together.



ABSTRACT

Cancer is listed in top ten global causes of death in 2016. The percentage of cancer incidence and mortality are steadily rising over the year. This necessitates alternate anti-cancer treatment as the conventional treatment have resulted in high number of long- term survivors who are left to deal with the aftereffect of their therapy. Drug delivery is a challenging aspect in medical community as the plasma membrane is non-permeable to external molecules. From this context, a study was proposed to use of natural plant extract *Artocarpus altilis* which will be uploaded into cervical cancer cell with the aid of electrical pulses or electroporation technique to investigate on the anti-cancer mechanism. Electroporation parameters utilized in this study ranging from 200V/cm, 400V/cm, 600V/cm, 800V/cm, 1000V/cm of pulse amplitude, while pulse duration ranging from 100 μ s, 500 μ s, 1ms, 5ms and 10ms with single pulse. Three parts of *artocarpus altilis* tested in this research study which is the peel, pulp and whole fruit. The concentration of all three parts which were tested on cervical cancer cell ranged from 10 μ g/ml, 20 μ g/ml, 30 μ g/ml, 40 μ g/ml, and 50 μ g/ml with methanol as the solvent. It is found that pulse electric field increased the cell spreading up to 129.8% with 600V/cm of pulse amplitude, which stimulates to the highest cell length percentage. Apart from that, cell viability found to be decrease with an increase in pulse amplitude and pulse duration. 600V/cm of pulse amplitude and 10ms of pulse duration stimulates highest cell viability with 53.3% as compared to control group (without treatment). Cell viability of cervical cancer cell measured when cell subjected to electrical field alone, extract alone and both electrical field and extract and the result of the study revealed that lowest cell viability stimulated by the combination of electrical pulse and extract up to 28% of cell viability. Thus the outcome of this research revealed new targets and electrochemical pathways for cervical cancer treatment while with the discovery health of patients will be maintained and there will be an improvement in general life expectancy.

ABSTRAK

Kanser disenaraikan dalam sepuluh penyebab utama kematian pada tahun 2016. Peratusan kejadian penyakit kanser dan kematian terus meningkat sepanjang tahun. Maka rawatan alternatif anti-kanser diperlukan kerana rawatan konvensional telah menyebabkan banyak kesan sampingan pada jangka masa yang panjang. Sistem penyampaian ubat merupakan aspek yang mencabar dalam bidang perubatan kerana membran plasma tidak menerima molekul luar. Dalam konteks ini, satu kajian dicadangkan untuk menggunakan ekstrak tumbuhan semulajadi Sukun dan bantuan medan elektrik denyut atau teknik electroporasi untuk menyiasat mekanisme anti kanser keatas sel kanser serviks. Parameter electroporasi yang digunakan dalam kajian ini terdiri daripada 200V/cm, 400V/cm, 600V/cm, 800V/cm, 1000V/cm amplitud denyut, manakala tempoh denyut adalah antara 100 μ s, 500 μ s, 1ms, 5ms dan 10ms. Tiga bahagian Sukun diuji dalam kajian penyelidikan ini iaitu kulit, pulpa dan buah tumbuhan tersebut. Konsentrasi ketiga-tiga bahagian yang diuji pada sel kanser serviks antara 10 μ g/ml, 20 μ g/ml, 30 μ g/ml, 40 μ g/ml, dan 50 μ g/ml dengan metanol sebagai pelarut. Adalah didapati bahawa medan elektrik denyut meningkatkan panjang sel sehingga 129.8% dengan amplitud nadi 600V/cm, yang merangsang peratusan panjang sel tertinggi. Selain itu, daya tahan sel didapati berkurangan dengan peningkatan amplitud nadi dan tempoh nadi. Amplitud denyut 600V/cm dan 10ms tempoh denyut merangsang daya maju sel terendah dengan peratusan sebanyak 53.3% berbanding dengan kumpulan sel kawalan. Daya tahan sel sel kanser serviks diukur apabila sel tertakluk kepada medan elektrik sahaja, ekstrak sahaja dan kedua-dua medan elektrik dan ekstrak dan hasil kajian mendedahkan bahawa daya tahan sel paling rendah dirangsang oleh gabungan nadi elektrik dan ekstrak sehingga 28% daripada daya tahan sel. Oleh itu, hasil kajian ini menunjukkan sasaran baru elektrokimia untuk rawatan kanser serviks malah kesihatan pesakit akan terjamin.

TABLE OF CONTENTS

TITLE	i
DECLARATION	ii
DEDICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	xii
LIST OF FIGURES	xiv
LIST OF SYMBOLS AND ABBREVIATION	xviii
LIST OF APPENDICES	xix
CHAPTER 1 INTRODUCTION	1
1.1 Background study	1
1.2 Problem statement	3
1.3 Objectives	4
1.4 Scope of study	4
1.5 Thesis structure	5
CHAPTER 2 LITERATURE REVIEW	6
2.1 Biological cells	6
2.2 Human cervical cancer cell	7
2.3 Composition of cell	7
2.3.1 Nucleus	8
2.3.2 Mitochondrion	9
2.3.3 Endoplasmic reticulum (ER)	10
2.3.4 Golgi apparatus	11
2.3.5 Cell membrane	12

2.4	Cell cycle	14
2.5	Apoptosis	15
2.6	Electroporation	16
2.6.1	Theory of electroporation	17
2.6.2	Types of electroporation	18
2.6.3	Parameters influencing electroporation	20
2.7	Anti-cancer studies	21
2.8	Artocarpus altilis overview	22
2.8.1	Taxonomy of artocarpus altilis	23
2.8.2	Botany description	23
2.8.3	Benefits of artocarpus altilis	24
2.8.4	Chemical constituents	25
CHAPTER 3	RESEARCH METHODOLOGY	26
3.1	Introduction	26
3.2	Material	29
3.2.1	Reagents	29
3.2.2	Equipment	34
3.3	Methods	40
3.3.1	Precautionary method	40
3.3.2	Cell culture protocol	41
3.3.3	Sub-culture protocol	42
3.3.4	Electroporation protocol	49
3.3.5	Cell counting method using Haemocytometer	50
3.3.6	Cell viability assay	52
3.3.7	Plant extraction	53
CHAPTER 4	PRELIMINARY STUDY ON CELL CULTURE AND ELECTROPORATION ON CERVICAL CANCER CELL	55
4.1	Introduction	55
4.2	Materials and method	56
4.2.1	Cell culture	56
4.3	Result and discussion	57



PTTA UTHM
PERPUSTAKAAN TUN AMINAH

4.3.1	Cell culture using HeLa cell	57
4.4	Electroporation effect on HeLa cell	62
4.5	Method	62
4.5.1	Cell culture and electroporation	62
4.6	Result on electroporation effect on HeLa cell	63
4.7	Discussion	65
4.8	Summary	66
CHAPTER 5	CORRELATION BETWEEN ELECTRICAL FIELD STRENGTH AND PULSE WIDTH ANALYSIS ON CELL VIABILITY	68
5.1	Introduction	68
5.2	Materials and methods	69
5.2.1	Cell culture	69
5.2.2	Electroporation	70
5.2.3	Measurement of cell viability	70
5.3	Result and discussion	72
5.3.1	HeLa cell viability	72
5.4	Summary	76
CHAPTER 6	ARTOCARPUS ALTILIS EXTRACT EFFECT ON HELA CANCER CELL	77
6.1	Breadfruit peel extract impact on hela cancer cell viability and proliferation	78
6.2	Materials and methods	78
6.2.1	Materials	78
6.2.2	Cell culture	78
6.2.3	Peel extract preparation	79
6.2.4	IC ₅₀ value determination	79
6.3	Result	80
6.4	Discussion	81
6.5	Influence of artocarpus altilis fruit extract on tumor cell	82
6.6	Influence of artocarpus altilis fruit extract on tumor cell	83



6.6.1	Materials	83
6.6.2	Cell culture	83
6.6.3	Whole fruit extract dilution	84
6.6.4	IC ₅₀ value determination	84
6.7	Result	85
6.8	Discussion	87
6.9	Pulp extract effect on cervical cancer cell	88
6.10	Materials and method	89
6.10.1	Materials	89
6.10.2	Cell culture	90
6.10.3	Pulp extract preparation	90
6.10.4	IC ₅₀ value determination	90
6.11	Result	91
6.12	Discussion	94
6.13	Summary	95
CHAPTER 7	INVESTIGATION OF ELECTROPORATION METHOD AND EXTRACT ON CERVICAL CANCER CELL	96
7.1	Electrical pulsed cervical cancer cell with whole fruit extract	96
7.2	Methods	97
7.2.1	Cell preparation	97
7.2.2	Extract dilution	98
7.2.3	IC ₅₀ value determination	98
7.2.4	Electroporation	98
7.3	Result	100
7.4	Discussion	103
7.5	Summary	104
7.6	Electrical pulsed cervical cancer cell with pulp part	104
7.7	Method	105
7.7.1	Cell preparation	105
7.7.2	Stock solution	106



7.7.3	Electroporation	106
7.8	Result	107
7.9	Discussion	110
7.10	Summary	111
7.11	Electrical pulsed cervical cancer cell with peel extract	111
7.12	Method	112
7.12.1	Cell preparation	112
7.12.2	Stock solution	112
7.12.3	Electroporation	113
7.13	Result	113
7.14	Discussion	116
7.15	Summary	117
CHAPTER 8	CONCLUSION	118
8.1	Conclusion	118
8.2	Recommendation	120
	REFERENCES	121
	APPENDICES	131
	VITA	138



LIST OF TABLES

2.1	Anti-cancer studies	21
2.2	Artocarpus altilis taxonomy	23
3.1	ECM 830 electroporator technical specification	38
3.2	Precautionary measures	40
3.3	Sub- culture process	42
3.4	Electroporation protocols	49
3.5	Working concentration	54
4.1	HeLa cell length	65
5.1	HeLa cell viability percentage with varied field strength	71
5.2	HeLa cell viability percentage with varied pulse duration	74
6.1	Cervical cell proliferation rate	80
6.2	Cervical cell viability with peel extract	80
6.3	Cell proliferation rate	86
6.4	Cell viability with whole fruit extract	86
6.5	HeLa cell viability with different concentration	92
6.6	HeLa cell viability with pulp extract	93
7.1	Pulse amplitude and actual value delivered	98
7.2	Pulse duration and actual value delivered	98
7.3	HeLa cell viability with varied pulse amplitude	101
7.4	HeLa cell viability with varied pulse duration	101
7.5	Extract concentration	105
7.6	HeLa cell viability with varied pulse amplitude	107
7.7	HeLa cell viability with varied pulse duration	108
7.8	Peel extract concentration	112

7.9	HeLa cell viability with varied pulse amplitude	113
7.10	HeLa cell viability with varied pulse duration	114



PTTA UTHM
PERPUSTAKAAN TUNKU TUN AMINAH

LIST OF FIGURES

2.1	Cervical cancer cell	7
2.2	Illustration of prokaryotic and eukaryotic Cells	8
2.3	Illustration of nucleus	9
2.4	Illustration of mitochondrion	10
2.5	Illustration of endoplasmic reticulum	11
2.6	Illustration of golgi apparatus	12
2.7	Cell membrane	13
2.8	Cell cycle process	14
2.9	Mitotic phase	15
2.10	Cytology of apoptosis	16
2.11	Electroporation process	17
2.12	Electrogenotherapy	19
2.13	Reversible and irreversible electroporation process	20
2.14	Relationship parameter between electric field and pulse duration time	20
2.15	The fruit of artocarpus altilis / breadfruit	24
2.16	Breadfruit with seed; Seedless breadfruit	24
3.1	Method of project represented by flow chart	27
3.2	RPMI 1640 media	29
3.3	Fetal bovine serum	30
3.4	Antibiotic	30
3.5	Phosphate buffer saline	31
3.6	Tryple express solution	32
3.7	Trypan blue	33
3.8	Biosafety Hood	34
3.9	Incubator	35

3.10	Water bath	35
3.11	Centrifuge	36
3.12	Refrigerator; Freezer	37
3.13	ECM 830 Electroporator	37
3.14	Nikon eclipse TS100 microscope	39
3.15	Haemocytometer and coverslip	51
3.16	Dilution of cell suspension with trypan blue solution	51
3.17	Chamber filled with diluted cell suspension	52
4.1	HeLa cells in 25cm ² flask at 80-90% confluence	57
4.2	Image of cell washing using PBS	58
4.3	Adding triple express solution	59
4.4	Image of cell after detachment	60
4.5	Growth of HeLa cell in 48 hours in 25cm ² flask	61
4.6	Electroporated and non electroporated cell after 48 hours in 25cm ² flask	63
4.7	Average Cell Length (µm) against Voltage (V/cm)	65
5.1	HeLa cell viability percentage with different field strength and fixed pulse width	72
5.2	Image of HeLa cell Electroporated with different field strength parameter	73
5.3	HeLa cell viability percentage with different pulse duration and fixed electric field strength of 600V/cm	74
6.1	Peel extract; Methanol; Stock solution	78
6.2	Serial dilution of artocarpus altilis peel extract	78
6.3	Photomicrograph of cervical cell line with different concentration of artocarpus altilis peel extract treatment	79
6.4	Cell proliferation against artocarpus altilis peel extract concentration	79
6.5	WF extract; Methanol; Stock solution	83
6.6	Serial Dilution of whole fruit of artocarpus altilis	83
6.7	Photomicrograph of HeLa cell with different concentration of WF treatment	84



6.8	Cell proliferation against Artocarpus altilis concentration	85
6.9	Stock solution	89
6.10	Serial dilution	89
6.11	Photomicrograph of HeLa cell with different concentration of treatment	90
6.12	Photomicrograph of HeLa cell with different concentration of treatment	91
6.13	Proliferation factor against pulp concentration	92
7.1	Image of HeLa cell under varied pulse amplitude, constant pulse duration and WF IC50 treatment	99
7.2	Image of HeLa cell under varied pulse duration, constant pulse amplitude and WF IC50 treatment	100
7.3	A graph of cell viability versus pulse amplitude	101
7.4	A graph of cell viability versus pulse duration	102
7.5	HeLa cell morphology under varied pulse amplitude, constant pulse duration and 30µg/ml pulp extract concentration	106
7.6	HeLa cell morphology under varied pulse duration, constant pulse amplitude and 30µg/ml pulp extract concentration	107
7.7	Cervical cancer cell viability with varied voltage, pulse duration at 200µs and 30µg/ml pulp extract concentration	108
7.8	Cervical cancer cell viability with varied pulse duration, voltage at 600V/cm and 30µg/ml pulp extract concentration	109
7.9	HeLa cell morphology under varied pulse amplitude, constant pulse duration and 39µg/ml peel extract concentration	112
7.10	HeLa cell morphology under varied pulse duration and constant pulse amplitude, 39µg/ml peel extract concentration	113



7.11	Cervical cancer cell viability against pulse amplitude and 39 μ g/ml peel extract concentration	114
7.12	Cervical cancer cell viability against pulse duration and 39 μ g/ml peel extract concentration	114



PTTA UTHM
PERPUSTAKAAN TUNKU TUN AMINAH

LIST OF SYMBOLS AND ABBREVIATIONS

<i>Cm</i>	Centimetre
<i>E</i>	Electric field strength
<i>ml</i>	Mililitre
<i>ms</i>	Milisecond
<i>nm</i>	Nanometre
<i>T</i>	Pulse duration
<i>V</i>	Voltage
μL	Microlitre
μs	Microsecond
CO_2	Carbon dioxide
$^{\circ}C$	Degree celsius
<i>DNA</i>	Deoxyribonucleic matrix
<i>ECM</i>	Extracellular matrix
<i>EF</i>	Electrofusion
<i>EG</i>	Electrogenetheraphy
<i>EP</i>	Electroporation
<i>ECT</i>	Electrochemotherapy
<i>FBS</i>	Fetal bovine serum
<i>HELA</i>	Cervical cancer cell
IC_{50}	Half maximal inhibitory concentration
<i>NEP</i>	Non-electroporation
<i>PBS</i>	Phosphate bovine serum
<i>RPMI</i>	Roswell Park Memorial Institute Media
<i>TES</i>	Tryple express solution

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Published journals and conference papers	131



PTTA UTHM
PERPUSTAKAAN TUNKU TUN AMINAH

LIST OF ASSOCIATED PUBLICATIONS

Journals

1. Ganeson, S., Jamil, M. M. A., Adon, M. N., Wahab, R. A., & Ahmad, M. S. (2017). Artocarpus altilis compound based study for anti-cancer proliferation on HeLa cell via Electroporation Method. *Journal of Fundamental and Applied Sciences*, 9(4S), 310-318. [ISI]
2. Jamil, M. M. A., Ganeson, S., Mammam, H. B. & Wahab, R. A. (2018). Artocarpus altilis extract effect on cervical cancer cells. *Materials Today: Proceedings*, 5(7), 15559-15566. [Scopus]
3. Ganeson, S., Jamil, M. M. A., Mamman, H. B., & Rahman, N. A. A. (2018). Correlation between electrical field strength and pulse width analysis on cell viability. In *Journal of Physics: Conference Series*, 1019(1), 012012. IOP Publishing. [ISI & Scopus]

Conference proceedings

1. Ganeson, S., Ambar, R., & Jamil, M. M. A. Influence of artocarpus altilis fruit extract on tumour cell. *2018 9th IEEE Control and System Graduate Research Colloquium (ICSGRC 2018)*. [Submitted]
2. Mahadi, M., Jamil, A., Mamman, H. B., & Ganeson, S. (2018, April). Investigation of Anti-Cancer Proliferation Properties of Luffa Acutangula on Ht29 Using Electroporation Method. In *Proceedings of the 2018 8th International Conference on Biomedical Engineering and Technology* (pp. 108-111). ACM.

Lecture Notes (Electrical Engineering LNEE-Springer)

1. Ganeson, S., Jamil, M. M. A., & Adon, M. N. Electrical pulsed artocarpus altilis on cervical cancer cell. *10th International Conference on Robotics, Vision, Signal Processing & Power Applications (ROVISP2018)*.
[Submitted]
2. Ganeson, S., Ambar, R., & Jamil, M. M. A. Breadfruit peel extract impact on hela cancer cell viability and proliferation. *10th International Conference on Robotics, Vision, Signal Processing & Power Applications (ROVISP2018)*.
[Submitted]



PTTA UTHM
PERPUSTAKAAN TUNKU TUN AMINAH

LIST OF COPYRIGHTS, AWARD AND SCHOLARSHIP

Copyrights

1. Cervical cancer cell inhibition via electroporation technique.
Registration Number: (LY2017005064) 2017.
2. Artocarpus altilis extract study on cervical cancer cell for anti-cancer treatment.
Registration number: (LY2017005066) 2017.

Award

1. **Faculty Level: Second Runner Up** in 3 Minutes Idea Presentation on 4 December 2016, Faculty of Electrical and Electronic Engineering.

Scholarship

1. Ministry of Higher Education, Malaysia. MyBrain15 (MyMaster).

International Product Innovation

1. Suhassni Ganeson, Muhammad Mahadi Abdul Jamil “Investigation of Electroporation Method and Extract on Cell Cervical Cancer Cell” Indonesia-Malaysia-Thailand Symposium on Innovation and Creativity (iMIT SIC 2018) Riau, Indonesia (August 2018). [Silver Medal]
2. Suhassni Ganeson, Muhammad Mahadi bin Abdul Jamil, Radzi bin Ambar, Ridhwan Abdul Wahab, Md Asri Ngadi, Johan Mohamad Sharif “Investigation of Electroporation Method and Extract on Cell Cervical Cancer

Cell” International Eureka Innovation Exhibition (i-EIE 2018) UniKL Kedah
(October 2018). [Bronze Medal]



CHAPTER 1

INTRODUCTION

1.1 Background of study

The number of cancer disease occurrences and mortality is expected to increase rapidly due to population growth, aging factor, and unhealthy lifestyle that increase the risk of cancer. Cancer is referred to an abnormal cells which replicates in uncontrolled manner in the body, accumulated different mutations and it occurs when the body's natural control mechanism fails to operate (Susan, 2010). Cancer can develop anywhere in the body and can be categorized based on either biological type or the initial site; part where the cancer originated. There are five major categories of cancer; sarcomas (develops in the bone, fat, muscles, cartilage, connective tissues), carcinoma (cancer that originated in the skin and tissues), leukemia (cancer that begins in blood-forming tissue), lymphomas (cancer that starts begins in the immune system) and myeloma (begins in the cell of immune system; bone marrow).

To date there are many treatments available in order to help the cancer patients, for an instance surgery, immune therapy, radiation therapy, hyperthermia and so on. Electrochemotherapy, the combination of electroporation and impermeant anti-cancer drugs, is one of the effective anti-cancer treatments. For a better understanding, it is an application of electroporation (short and intense electric pulses) which increases cell membrane permeability, and enhance the uptake of anti-cancer drugs to treat the tumorous cells. Cisplatin and bleomycin have proven to be the most appropriate drugs to be use with electroporation due to the chemical properties.

However, advances in cancer treatment have resulted in high number of long- term survivors who were left to deal with the aftereffect of their treatments. Drugs usage in cancer therapy leads to multiple permanent side effects. The use of bleomycin drugs associated with fatal side effects like induction of pulmonary toxicity. Utilization of bleomycin, will cause pulmonary syndromes such as bronchiolitis obliterans, eosinophilic hypersensitivity and interstitial pneumonitis (Sleijfer, 2001). On the other hand, neurotoxicity, nephrotoxicity, ototoxicity, nausea and vomiting are the side effects of cisplatin drug. Moreover this type of drugs has the potential in interrupting treatments with effectiveness reduction and affect patient's quality of life (Santabarbara, Maione, Rossi, & Gridelli, 2016). Apart from anti-cancer drugs, patients undergoing electrochemotherapy felt of discomfort due to muscle contraction during to the high frequency electric pulses delivery (Mamman, Jamil, & Adon, 2017).

There are an increasing number of new promising studies of anti-cancer using natural plants as chemotherapeutic agent. Natural plants that contain anti-cancer properties like Curcumin and Artocarpus altilis wood which inhibit proliferation in breast cancer cell. Apart from that, jackfruits have also shown anti-cancer properties against laryngeal, cervical, breast and lung cancer. Moreover, bioactive compounds are found in various types of berries and saffron proved to have chemo-preventive properties. Recent studies claimed that the combination of turmeric, ginger and garlic established to oppose the growth of breast cancer cell.

This study focuses on Artocarpus altilis to investigate the anti-proliferation on cervical cancer cell. Artocarpus altilis is an extensively grown and nutritious tree fruit. It is also known as breadfruit while *Sukun* in Malaysia and Artocarpus altilis scientifically. Each of every parts of Artocarpus altilis has its own importance and it is widely distributed in tropical and subtropical regions including Malaysia and Indonesia (Fajriah, Mozef, Artanti, Lotulung, & Abbas, 2013). There are numbers of studies focusing on Artocarpus altilis, and proven to have several interesting biological properties including anti-cancer, antioxidant, antibacterial, anti-inflammatory, antihypertensive and many more (Fajriah et al., 2013; Vasugi, Sudhahar, & Anandajogopal, 2012). As the use of drugs in cancer therapies, comes with multiple side effects, Artocarpus altilis has been chosen for this study taking into consideration its anti-cancer properties to inhibit cancer cell proliferation.

Since the Artocarpus altilis extract is impermeant, or in another word unable

REFERENCES

- Acton, R. (2012). *Cell culture and its application*: Elsevier.
- Aggarwal, B. B., Ichikawa, H., Garodia, P., Weerasinghe, P., Sethi, G., Bhatt, I. D., & Nair, M. G. (2006). From traditional Ayurvedic medicine to modern medicine: identification of therapeutic targets for suppression of inflammation and cancer. *Expert Opinion on Therapeutic Targets*, 10(1), 87-118.
- Aggarwal, B. B., Surh, Y. J., & Shishodia, S. (2007). *The molecular targets and therapeutic uses of curcumin in health and disease* (Vol. 595). Springer Science & Business Media.
- Ahamad, M. S., Siddiqui, S., Jafri, A., Ahmad, S., Afzal, M., & Arshad, M. (2014). Induction of apoptosis and antiproliferative activity of naringenin in human epidermoid carcinoma cell through ROS generation and cell cycle arrest. *PloS One*, 9(10), e110003.
- Akbar, E., Yaakob, Z., Kamarudin, S. K., Ismail, M., & Salimon, J. (2009). Characteristic and composition of Jatropha curcas oil seed from Malaysia and its potential as biodiesel feedstock feedstock. *European Journal of Scientific Research*, 29(3), 396-403.
- Arung, E. T., Wicaksono, B. D., Handoko, Y. A., Kusuma, I. W., Yulia, D., & Sandra, F. (2009). Anti-cancer properties of diethylether extract of wood from sukun (*Artocarpus altilis*) in human breast cancer (T47D) cells. *Tropical Journal of Pharmaceutical Research*, 8(4).
- Asadi-Samani, M., Kooti, W., Aslani, E., & Shirzad, H. (2016). A systematic review of Iran's medicinal plants with anticancer effects. *Journal of Evidence-based Complementary & Alternative Medicine*, 21(2), 143-153.
- B. Amirlak, L. S., S. Javaheri, F. Talavera, and W. Stadelmann. (2011). Skin anatomy. *emedicine medspace_article*.
- Bertacchini, C., Margotti, P. M., Bergamini, E., Lodi, A., Ronchetti, M., & Cadossi,

- R. (2007). Design of an irreversible electroporation system for clinical use. *Technology in Cancer Research & Treatment*, 6(4), 313-320.
- Bharti, A. C., Donato, N., & Aggarwal, B. B. (2003). Curcumin (diferuloylmethane) inhibits constitutive and IL-6-inducible STAT3 phosphorylation in human multiple myeloma cells. *The Journal of Immunology*, 171(7), 3863-3871.
- Bianconi, E., Piovesan, A., Facchin, F., Beraudi, A., Casadei, R., Frabetti, F., Vitale, L., Pelleri, M. C., Tassani, S., Piva, F., & Perez-Amodio, S. (2013). An estimation of the number of cells in the human body. *Annals of Human Biology*, 40(6), 463-471.
- Carter, M., & Shieh, J. C. (2015). *Guide to research techniques in neuroscience*. Academic Press.
- Castillo-Pichardo, L., Martínez-Montemayor, M. M., Martínez, J. E., Wall, K. M., Cubano, L. A., & Dharmawardhane, S. (2009). Inhibition of mammary tumor growth and metastases to bone and liver by dietary grape polyphenols. *Clinical & Experimental Metastasis*, 26(6), 505-516.
- Cemazar, M., Ambrozic Avgustin, J., Pavlin, D., Sersa, G., Poli, A., Krhac Levacic, A., & Tozon, N. (2017). Efficacy and safety of electrochemotherapy combined with peritumoral IL-12 gene electrotransfer of canine mast cell tumours. *Veterinary and Comparative Oncology*, 15(2), 641-654.
- Charrier-Savournin, F. B., Château, M. T., Gire, V., Sedivy, J., Piette, J., & Dulić, V. (2004). p21-Mediated nuclear retention of cyclin B1-Cdk1 in response to genotoxic stress. *Molecular Biology of the Cell*, 15(9), 3965-3976.
- Chu, G., Hayakawa, H., & Berg, P. (1987). Electroporation for the efficient transfection of mammalian cells with DNA. *Nucleic Acids Research*, 15(3), 1311-1326.
- Cooper, G. M. (2000). The eukaryotic cell cycle. *The Cell: A Molecular Approach* (2nd).
- Davidson, M. W. (1995-2017). Molecular Expression ; Golgi Apparatus.
- Deivanai, S., & Subhash, J. B. (2010). Breadfruit (*Artocarpus altilis* Fosb.)—An underutilized and neglected fruit plant species. *Middle-East Journal of Scientific Research*, 6(5), 418-428.
- Donaldson, C. D., & Bishop, K. N. (2015). Cell culture. *British Journal of Hospital Medicine*, 76(1), C2-C5. doi: <https://doi.org/10.12968/hmed.2015.76.1.C2>
- Fajriah, S., Mozef, T., Artanti, N., Lotulung, P. D., & Abbas, J. (2013). Isolation of

prenylated flavonoid from ethyl acetate fraction of artocarpus altilis leaves using counter-current chromatography. *Asian Transactions on Basic and Applied Sciences*, 2(06), 6-9.

Faria, A., Calhau, C., de Freitas, V., & Mateus, N. (2006). Procyanidins as antioxidants and tumor cell growth modulators. *Journal of Agricultural and Food Chemistry*, 54(6), 2392-2397.

Fresco, P., Borges, F., Diniz, C., & Marques, M. P. M. (2006). New insights on the anticancer properties of dietary polyphenols. *Medicinal Research Reviews*, 26(6), 747-766.

Gothelf, A., Mir, L. M., & Gehl, J. (2003). Electrochemotherapy: results of cancer treatment using enhanced delivery of bleomycin by electroporation. *Cancer Treatment Reviews*, 29(5), 371-387.

Granot, Y., & Rubinsky, B. (2008). Mass transfer model for drug delivery in tissue cells with reversible electroporation. *International Journal of Heat and Mass Transfer*, 51(23-24), 5610-5616.

Greenwell, M., & Rahman, P. K. S. M. (2015). Medicinal plants: their use in anticancer treatment. *International Journal of Pharmaceutical Sciences and Research*, 6(10), 4103.

Heller, R., Jaroszeski, M. J., Glass, L. F., Messina, J. L., Rapaport, D. P., DeConti, R. C., Fenske, N. A., Gilbert, R. A., Mir, L. M., & Reintgen, D. S. (1996). Phase I/II trial for the treatment of cutaneous and subcutaneous tumors using electrochemotherapy. *Cancer: Interdisciplinary International Journal of the American Cancer Society*, 77(5), 964-971.

HPV Information Centre. (2017). *Human Papillomavirus and Related Diseases Report*. Barcelona: HPV Information Centre.

Hui, S. W. (1995). Effects of pulse length and strength on electroporation efficiency. *Plant Cell Electroporation and Electrofusion Protocols*, 29-40.

International Agency for Research on Cancer. (2012). Cervical cancer estimated incidence, mortality and prevalence worldwide in 2012. *World Health Organization*.

Jagtap, U. B., & Bapat, V. A. (2010). Artocarpus: A review of its traditional uses, phytochemistry and pharmacology. *Journal of Ethnopharmacology*, 129(2), 142-166.

Jalal, T. K., Ahmed, I. A., Mikail, M., Momand, L., Draman, S., Isa, M. L. M.,

- Rasad, M. S. B. A., Omar, M. N., Ibrahim, M. & Wahab, R. A. (2015). Evaluation of antioxidant, total phenol and flavonoid content and antimicrobial activities of *Artocarpus altilis* (breadfruit) of underutilized tropical fruit extracts. *Applied Biochemistry and Biotechnology*, 175(7), 3231-3243.
- Jaroszeski, M. J., Gilbert, R., & Heller, R. (1997). Electrochemotherapy: an emerging drug delivery method for the treatment of cancer. *Advanced Drug Delivery Reviews*, 26(2-3), 185-197.
- Jeon, Y. J., Jung, S. N., Chang, H., Yun, J., Lee, C. W., Lee, J., Choi, S., Nash, O., Han, D. C., & Kwon, B. M. (2015). *Artocarpus altilis* (Parkinson) Fosberg extracts and geranyl dihydrochalcone inhibit STAT3 activity in prostate cancer DU145 cells. *Phytotherapy Research*, 29(5), 749-756.
- Johnson, L. B. (2005). Electroporation System for Sterilizing Water. *Article Tech Brief*. Retrieved from <https://www.techbriefs.com/component/content/article/tb/techbriefs/bio-medical/271>.
- Kinghorn, A. D. (2015). Review of anticancer agents from natural products. *Journal of Natural Products*, 78(9), 2315-2315.
- Korohoda, W., Grys, M., & Madeja, Z. (2013). Reversible and irreversible electroporation of cell suspensions flowing through a localized DC electric field. *Cellular & Molecular Biology Letters*, 18(1), 102.
- Kuwajerwala, N., Cifuentes, E., Gautam, S., Menon, M., Barrack, E. R., & Reddy, G. P. V. (2002). Resveratrol induces prostate cancer cell entry into s phase and inhibits DNA synthesis. *Cancer Research*, 62(9), 2488-2492.
- Larkin, J. O., Collins, C. G., Aarons, S., Tangney, M., Whelan, M., O'reily, S., Breathnach, O., Soden, D. M., and O'sullivan, G. C. (2007). Electrochemotherapy: aspects of preclinical development and early clinical experience. *Annals of Surgery*, 245(3), 469.
- Lenyo, J., Perez, C., & Owens, H. Cell Membrane (2015).
- Lippman, S. M., Lee, J. J., & Sabichi, A. L. (1998). Cancer chemoprevention: progress and promise. *Journal of the National Cancer Institute*, 90(20), 1514-1528.
- Lloyd, A. C. (2013). The regulation of cell size. *Cell*, 154(6), 1194-1205.
- Love, S. M. (2010). *Dr. Susan Love's Breast Book, (A Merloyd Lawrence Book)*. Da Capo Lifelong Book.

- Mahnič-Kalamiza, S., Vorobiev, E., & Miklavčič, D. (2014). Electroporation in food processing and biorefinery. *The Journal of membrane biology*, 247(12), 1279-1304.
- Mamman, H. B., Jamil, M. M. A., & Adon, M. N. (2016). Optimization of electric field parameters for HT29 cell line towards wound healing application. *Indian Journal of Science and Technology*, 9(46), 1-6.
- Mamman, H. B., Jamil, M. M. A., & Adon, M. N. (2017). Low amplitude pulse electric field for elimination of unpleasant sensation associated with high amplitude electric field for electrochemotherapy. *Journal of Advances in Technology and Engineering Studies*, 3(2), 27-33.
- Mamman, H. B., Sadiq, A., Adon, M. N., & Jamil, M. M. A. (2015). Study of electroporation effect on HT29 cell migration properties. In *Control System, Computing and Engineering (ICCSCE), 2015 IEEE International Conference*. IEEE. 342-346.
- Manabe, Y., Nakagawa, R., Zhehong, S., Maetani, M., Teranishi, K., Shimomura, N., & Takahashi, A. (2011). Influences of pulsed electric fields on the gene expression of pathogenic bacteria. In *Pulsed Power Conference (PPC)*. IEEE. 1242-1246.
- Mann, J. (2002). Natural products in cancer chemotherapy: past, present and future. *Nature Reviews Cancer*, 2(2), 143.
- Miller, K. D., Siegel, R. L., Lin, C. C., Mariotto, A. B., Kramer, J. L., Rowland, J. H., Stein, K. D., Alteri, R., & Jemal, A. (2016). Cancer treatment and survivorship statistics, 2016. *CA: A Cancer Journal for Clinicians*, 66(4), 271-289.
- Minuth, W. W., Strehl, R., & Schumacher, K. (2005). *Tissue engineering: from cell biology to artificial organs*. Wiley-Blackwell.
- Mittal, L., Raman, V., Camarillo, I. G., & Sundararajan, R. (2017). Ultra-microsecond pulsed curcumin for effective treatment of triple negative breast cancers. *Biochemical and Biophysical Research Communications*, 491(4), 1015-1020.
- Mohanty, M., & Pradhan, C. (2015). A review on phytochemistry, bio-efficacy, medicinal and ethno-pharmaceutical importance of *Artocarpus altilis*. *International Journal of Pharmacy and Pharmaceutical Research*, 3, 219-231.

- Moreira, A. R. (2007). The evolution of protein expression and cell culture. *Biopharm International*, 20(10).
- Mukhopadhyay, A., Bueso-Ramos, C., Chatterjee, D., Pantazis, P., & Aggarwal, B. B. (2001). Curcumin downregulates cell survival mechanisms in human prostate cancer cell lines. *Oncogene*, 20(52), 7597.
- Munoz, N., Bosch, F. X., & Jensen, O. M. (1989). Human papillomavirus and cervical cancer. *IARC Scientific Publication*, 94.
- Neu, W. K., & Neu, J. C. (2009). Theory of electroporation. *Cardiac Bioelectric Therapy*, 133-161.
- Newman, D. J., & Cragg, G. M. (2016). Natural products as sources of new drugs from 1981 to 2014. *Journal of Natural Products*, 79(3), 629-661.
- Nguyen, M. T. T., Nguyen, N. T., Nguyen, K. D. H., Dau, H. T. T., Nguyen, H. X., Dang, P. H., Le, T. M., Phan, T. H. N., Tran, A. H., Nguyen, B. D. & Ueda, J. Y. (2014). Geranyl dihydrochalcones from *Artocarpus altilis* and their antiausteric activity. *Planta Medica*, 80(02/03), 193-200.
- Nickoloff, J. A. (Ed.) (1995). *Electroporation protocols for microorganisms*. Springer Science & Business Media, 47.
- Nomura, T., Hano, Y., & Aida, M. (1998). Isoprenoid-substituted flavonoids from *Artocarpus* plants (Moraceae). *Heterocycles*, 2(47), 1179-1205.
- Oeffinger, K. C., Fontham, E. T., Etzioni, R., Herzig, A., Michaelson, J. S., Shih, Y. C. T., Walter, L. C., Church, T. R., Flowers, C. R., LaMonte, S. J., & Wolf, A. M. (2015). Breast cancer screening for women at average risk: 2015 guideline update from the American Cancer Society. *Jama*, 314(15), 1599-1614.
- Orwa, M., Kindt, Jamnadass, Anthony (2009). *Artocapus Altilis*. Agroforestry Database.
- Parajuli, S., Pun, N. T., Parajuli, S., & Jamarkattel-Pandit, N. (2012). Antioxidant activity, total phenol and flavonoid contents in some selected medicinal plants of Nepal. *JHAS*, 2(1), 27-31.
- Park, H. S., Hwang, H. J., Kim, G. Y., Cha, H. J., Kim, W. J., Kim, N. D., Yoo, Y. H., & Choi, Y. H. (2013). Induction of apoptosis by fucoidan in human leukemia U937 cells through activation of p38 MAPK and modulation of Bcl-2 family. *Marine Drugs*, 11(7), 2347-2364.
- Patel, R. M., & Patel, S. K. (2011). Cytotoxic activity of methanolic extract of



- Artocarpus heterophyllus against A549, Hela and MCF-7 cell lines. *Journal of Applied Pharmaceutical Science*, 1(7), 167-171.
- Plotkin, S. (2014). History of vaccination. *Proceedings of the National Academy of Sciences*, 111(34), 12283-12287.
- Posakony, J. W., England, J. M., & Attardi, G. (1977). Mitochondrial growth and division during the cell cycle in HeLa cells. *The Journal of Cell Biology*, 74(2), 468-491.
- Pradhan, C., & Mohanty, M. (2014). Phytoconstituent analysis and comparative bioefficacy assessment of breadfruit leaf and fruit extracts for antipathogenic potentiality. *American Journal of Phytomedicine and Clinical Therapeutics*, 2(1), 77-87.
- Pradhan, C., Mohanty, M., & Rout, A. (2012). Phytochemical screening and comparative bioefficacy assessment of Artocarpus altilis leaf extracts for antimicrobial activity. *Frontiers in Life Science*, 6(3-4), 71-76.
- Pradhan, C., Mohanty, M., & Rout, A. (2013). Assessment of the antibacterial potential of breadfruit leaf extracts against pathogenic bacteria. *International Journal of Pharmaceutics*, 3, 374-379.
- Pradhan, C., Mohanty, M., Rout, A., Das, A. B., Satapathy, K. B., & Patra, H. K. (2013). Phytoconstituent screening and comparative assessment of antimicrobial potentiality of Artocarpus altilis fruit extracts. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(3), 840-843.
- Ragone, D. (1997). *Breadfruit: Artocarpus altilis (Parkinson) Fosberg-Promoting the conservation and use of underutilized and neglected crops*. 10. Bioversity International.
- Rajendran, N. K., & Jayapradha, R. (2010). Polyphenol analysis and anti-tumor activity of crude extracts from tegmen of Artocarpus heterophyllus. *Medicinal Plants*, 2(1), 63-66.
- Rems, L., & Miklavčič, D. (2016). Tutorial: electroporation of cells in complex materials and tissue. *Journal of Applied Physics*, 119(20), 201101.
- Rhoads, D. (2007). History of Cell Biology.
- Riggio, G. (2017). 5 Stages of Mitosis.
- Rodamporn, S. (2012). Optimal parameters of electroporation for gene and tissue. In *Biomedical Engineering International Conference (BMEiCON), 2011*. IEEE. 279-282.



PTTA (Pusat Teknologi dan Terapi) is a research center under the supervision of the Ministry of Health of the Republic of Indonesia. The center is located in the campus of Padjadjaran University (UNPAD) in Bandung. The center is dedicated to research and development of new drugs and medical devices. The center is also involved in the training of health professionals and the dissemination of research results.

- Rogers, K. (2017). Endoplasmic reticulum (ER). *Encyclopedia Britannica*.
- Saleem, A., Husheem, M., Härkönen, P., & Pihlaja, K. (2002). Inhibition of cancer cell growth by crude extract and the phenolics of *Terminalia chebula* retz. fruit. *Journal of Ethnopharmacology*, 81(3), 327-336.
- Sampath, B., Sivakumar, V., Sankaranarayanan, K., Camarillo, I. G., Lin, W. Y., Natarajan, A., & Sundararajan, R. (2014). Breast cancer treatment using curcumin, the natural herbal biodielectric. In *Electrical Insulation and Dielectric Phenomena (CEIDP), 2014 IEEE Conference*. IEEE. 235-238.
- Santabarbara, G., Maione, P., Rossi, A., & Gridelli, C. (2016). Pharmacotherapeutic options for treating adverse effects of Cisplatin chemotherapy. *Expert Opinion on Pharmacotherapy*, 17(4), 561-570.
- Saslow, D., Solomon, D., Lawson, H. W., Killackey, M., Kulasingam, S. L., Cain, J., Garcia, F. A., Moriaty, A. T., Waxman, A. G., Wilbur, D. C., & Wentzensen, N. (2012). American Cancer Society, American Society for Colposcopy and Cervical Pathology, and American Society for Clinical Pathology screening guidelines for the prevention and early detection of cervical cancer. *CA: A Cancer Journal for Clinicians*, 62(3), 147-172.
- Schlager, K. J. (2001). An electroporation drinking water disinfection system. *Final Report*. United States Environmental Protection Agency.
- Senawong, T., Khaopha, S., Misunaa, S., Komaikula, J., Senawonga, G., Wongphakhama, P., & Yunchalard, S. (2014). Phenolic acid composition and anticancer activity against human cancer cell lines of the commercially available fermentation products of *Houttuynia cordata*. *Sci Asia*, 40, 420-427.
- Shafri, M. A. M., Yusof, F. A., & Zain, A. Z. M. (2015). In vitro cytotoxic activity of *ferula assafoetida* on osteosarcoma cell line (HOS CRL). *Jurnal Teknologi*, 77(3).
- Sikarwar, M. S., Hui, B. J., Subramaniam, K., Valeisamy, B. D., Yean, L. K., & Balaji, K. (2014). A review on *Artocarpus altilis* (Parkinson) Fosberg (breadfruit). *Journal of Applied Pharmaceutical Science*, 4(8), 91-97.
- Siwak, D. R., Shishodia, S., Aggarwal, B. B., & Kurzrock, R. (2005). Curcumin-induced antiproliferative and proapoptotic effects in melanoma cells are associated with suppression of kinase and nuclear factor B activity and are independent of the BRAf mitogen activated extracellular signal regulated protein kinase pathway and the Akt pathway. *Cancer*, 104(4), 879-890.



- Skloot, R. (2017). *The immortal life of Henrietta Lacks*. Broadway Books.
- Sleijfer, S. (2001). Bleomycin-induced pneumonitis. *Chest Journal*, 120(2), 617-624.
- Somashekhar, M., Nayeem, N., & Sonnad, B. (2013). A review on family Moraceae (Mulberry) with a focus on Artocarpus species. *World Journal of Pharmacy and Pharmaceutical Sciences*, 2, 2614-2621.
- Stewart, B., & Wild, C. P. (2017). World cancer report 2014. *Report*. World Health Organization, International Agency for Research on Cancer.
- Sundararajan, R., Xiao, F., Salameh, T., Reece, L. M., Leary, J. F., Otto, K., Campana, L. G., & Campana, L. G. (2012). Effective proliferation control of human cancer cells using electrical pulses. *IEEE Transactions on Dielectrics and Electrical Insulation*, 19(6).
- Tarek, M. (2005). Membrane electroporation: a molecular dynamics simulation. *Biophysical Journal*, 88(6), 4045-4053.
- Tian, Q., & Zang, Y. H. (2015). Antiproliferative and apoptotic effects of the ethanolic herbal extract of *Achillea falcata* in human cervical cancer cells are mediated via cell cycle arrest and mitochondrial membrane potential loss. *Journal of the Balkan Union of Oncology*, 20(6), 1487-1496.
- Torre, L. A., Bray, F., Siegel, R. L., Ferlay, J., Lortet-Tieulent, J., & Jemal, A. (2015). Global cancer statistics, 2012. *CA: A Cancer Journal for Clinicians*, 65(2), 87-108.
- Tryfona, T., & Bustard, M. T. (2008). Impact of pulsed electric fields on *Corynebacterium glutamicum* cell membrane permeabilization. *Journal of Bioscience and Bioengineering*, 105(4), 375-382.
- Tsong, T. Y. (1991). Electroporation of cell membranes. *Biophysical Journal*, 60(2), 297-306.
- Vasugi, R., Sudhahar, D., & Anandajogopal, K. (2012). Preliminary phytochemical investigation and screening of antimicrobial activity of leaf extracts of *Artocarpus altilis*. *Asian Journal of Biological Life Sciences*, 4, 105-106.
- Vemuri, S. K., Banala, R. R., Subbaiah, G. P. V., Srivastava, S. K., Reddy, A. G., & Malarvili, T. (2017). Anti-cancer potential of a mix of natural extracts of turmeric, ginger and garlic: A cell-based study. *Egyptian Journal of Basic and Applied Sciences*, 4(4), 332-344.
- Wang, Y., Deng, T., Lin, L., Pan, Y., & Zheng, X. (2006). Bioassay guided isolation of antiatherosclerotic phytochemicals from *Artocarpus altilis*. *Phytotherapy*



- Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 20(12), 1052-1055.
- Wang, Y., Xu, K., Lin, L., Pan, Y., & Zheng, X. (2007). Geranyl flavonoids from the leaves of *Artocarpus altilis*. *Phytochemistry*, 68(9), 1300-1306.
- Weaver, J. C. (2000). Electroporation of cells and tissues. *IEEE Transactions on Plasma Science*, 28(1), 24-33.
- Wenzel, U., Kuntz, S., Brendel, M. D., & Daniel, H. (2000). Dietary flavone is a potent apoptosis inducer in human colon carcinoma cells. *Cancer Research*, 60(14), 3823-3831.
- Yarmush, M. L., Golberg, A., Serša, G., Kotnik, T., & Miklavčič, D. (2014). Electroporation-based technologies for medicine: principles, applications, and challenges. *Annual Review of Biomedical Engineering*, 16, 295-320.
- Yin, M. C., Lin, C. C., Wu, H. C., Tsao, S. M., & Hsu, C. K. (2009). Apoptotic effects of protocatechuic acid in human breast, lung, liver, cervix, and prostate cancer cells: potential mechanisms of action. *Journal of Agricultural and Food Chemistry*, 57(14), 6468-6473.
- Zaltum, M. A. M., Adon, M. N., & Jamil, M. M. A. (2013). Electroporation effect on growth of HeLa cells. In *The 6th 2013 Biomedical Engineering International Conference (BMEiCON), 2013*. IEEE. 1-4.
- Zegman, Y., Bonazzi, D., & Minc, N. (2015). Measurement and manipulation of cell size parameters in fission yeast. *Methods in Cell Biology*, 125, 423-436.

